



CODEN [USA]: IAJPBB

ISSN : 2349-7750

**INDO AMERICAN JOURNAL OF
PHARMACEUTICAL SCIENCES**

SJIF Impact Factor: 7.187

<https://doi.org/10.5281/zenodo.5595138>Available online at: <http://www.iajps.com>

Research Article

**SECONDARY METABOLITES AND THEIR BIOACTIVE
POTENTIAL ANALYZED BY GC-MS IN *STROBILANTHES
ALTERNATA* (BURM.F.) MOYLAN EX J.R.I.WOOD****Karthika Devarajan and *Hussain, K.**

Department of P.G and Research in BotanySNGS College, Pattambi, 679306- Kerala

Article Received: September 2021 **Accepted:** September 2021 **Published:** October 2021**Abstract:**

Bioactive secondary metabolites in the leaf extracts of Strobilanthes alternata were analysed using GCMS. The alcoholic extract of the leaves contain eight fatty acid methyl esters, five terpenoids and aromatic dicarboxylic acids distributed in varying concentrations. Squalene which occurred very low quantity compared to other molecules is reported to possess antioxidant potential and contribute considerable to the wound healing property. Most of the molecules are known to possess antibacterial, antifungal and anti-inflammatory activities. Many of the biomolecules are characterized by antioxidant potential also. The process of wound healing in which S.alternata known to play a vital role which is discussed in terms of the combined activity of each and every molecules of the leaf extracts possessing antimicrobial, anti-inflammatory and antioxidant activities. Wound healing and antioxidant potential of bioactive molecules are seemed to coexist in S.alternata.

Key Words: *Strobilanthes alternata, GCMS, Antioxidant, wound healing, Antimicrobial*

Corresponding author:**Hussain, K,**

Assistant Professor, SNGS College,

Pattambi, Palakkad. Kerala .

hussainkoorimannil@gmail.com

Ph: 6282273296

QR code



Please cite this article in press Karthika Devarajan and *Hussain, K, *Secondary Metabolites and their Bioactive Potential Analyzed by GC-MS in Strobilanthes Alternata (BURM.F.) Moylan Ex J.R.I.Wood., Indo Am. J. P. Sci, 2021; 08(10).*

1. INTRODUCTION:

More than four billion population of the world, that is about 80% of the world population presently use the traditional system of medicines or herbal drugs for primary health care. In India more than 65% of the total population use herbal medicinal products for the treatment of diseases [1].

For centuries, plants have been used to treat diseases worldwide and still performing a major role in health care system in many developing countries. The use of plant extracts and plant derived components in the treatment and management of wounds have been well observed [2,3,4,5].

Analysis of bioactive components of green plants leaves using GC-MS studies have been conducted in many medicinal plants. Important aspect repeatedly reported by these investigation is the phytochemical activities or potential of the extracts aqueous/organic solvents on microbes or on test animals . GC-MS analysis of *Baccopa monerrii* revealed qualitative and quantitative distribution of saponins[6] . Singh and Aeri ,(2013) analyzed secondary metabolites of *Acanthes ilicifolius* and reported components like saponins, tannins, alkaloids, flavonoids and triterpenoids[7]. *Croton bonplanbianum* plants revealed the occurrence of many secondary components possessing antibacterial, antitubercular and anticytotoxic activity by GC-MS studies[8]. According to Abid and Touquer (2015) phytochemicals of *Breynia disticha* include antimicrobial and antioxidant activities in methanolic extracts[9]. Tamboli *et al.*,(2018) compared the presence of phytoconstituents like alkaloids, phenolics, saponins and pyrenoids in natural and micropropagated from *Bacopa* plants[10].

Almost all studies using GC-MS to analyse bioactive components are centred around the identification and estimation of these metabolites of plants in general and medicinal plants in particular. GC- MS analyses of phytochemicals and potentially bioactive components of many medicinal plants have been reported in plants like *Euphorbia kansuii* [11], *Cassia italic*[12] , *Artimesia princeps* [13], *Moringa peregram* [14], *Vernonia elangifolia* and *Bregaria disticha* [9], *Moringa olifera* [15], *Huru crepitans* [16], *Ipomea violaeae* [17], *Equisetum arvanse* [18], *Bacopa monnerri* [10], *Homolomena aromatic* [19].

Antibacterial, antifungal, antioxidant and/or anti-inflammatory activities of several medicinal plants inclusive of *S.alternata* have been elaborately studied

and reported. Excision wound healing and anti-inflammatory properties of *Hemigraphis colorata* leaves were reported earlier [20,21,22]. Antibacterial activity of the plant was investigated against selected pathogens [23]. Antimicrobial, anti-inflammatory and antioxidant properties of *H.colorata* leaf extract was also reported [24]. Nevertheless, characteristic potential and activities of each component of the extracts of *S.alternata* have not yet been studied in detail.

In the present study, the qualitative and functional role of individual bioactive component separated by GC-MS in the methanolic extract of *S.alternata* (Syn: *Hemigraphis colorata*) leaves are compared and discussed in relation to the specific phytochemical characters or activities of each component molecules. Comparison is also made with the reported/attribution medicinal properties of the plant extracts irrespective of the solvent system used. *Strobilanthes alternata* (Burm.f.) E. Moylan ex J.R.I. wood (syn: *Hemigraphis colorata*) belongs to the family Acanthaceae. In kerala, the plant is popular in the name "Murikootti" and "Murianpacha", due to its incredible potency to heal fresh wounds [20]. It is prostrate plant spreads with rooting stem. Leaves are entire, lences shaped, toothed or lobed margin. Leaves of *S.alternata* are used for wound- healing purposes.

Keeping in view the antimicrobial and anti-inflammatory properties and other medicinal uses reported earlier, *S.alternata* is to be analyzed by GC-MS to evaluate the function of each and every bioactive components. Hence the present study was undertaken to separate and identify the secondary metabolites by GC- MS method to assess the function or vital role of the constituents of the leaf extract. The significance of leaf components in the processes of wound healing partaning to the antimicrobial and anti-inflammatory activities with more emphasis on antioxidant potential of specific bioactive molecules of *S.alternata* is the prime objective of this investigation.

2. MATERIALS AND METHODS:

2.1 Choice of plant

Mature leaves of healthy twigs of *Strobilanthes alternata* plants growing in the botanical garden, SNGS College Pattambi were collected and used for the study.

2.2 Preparation of leaf extract

Fresh leaves were collected and shade dried. Dried leaves were grounded into fine powder and 5g of the powder was subjected to extraction using 100ml of

methanol in soxhlet apparatus. After running several cycles, the extracts obtained was concentrated and was used for GC-MS analysis.

2.3 GC-MS Analysis

Gas Chromatography Mass Spectrometry analysis of leaf extracts was performed using Shimadzu GC-MS, with model number QP2010s, at Kerala Forest Research Institute, Thrissur, Kerala. Helium gas was used as the carrier gas at constant flow rate of 1ml/min and an injection volume of 1 μ L was employed. The injection port temperature was set at 260°C and ion source temperature at 200°C. The oven temperature was programmed from 80°C for 4 minutes with an increase of 5°C/min to 280°C with a hold time of 6 minutes. The components were identified based on the comparison of their relative

retention time and mass spectra with those of wiley NIST7Nlibrary data.

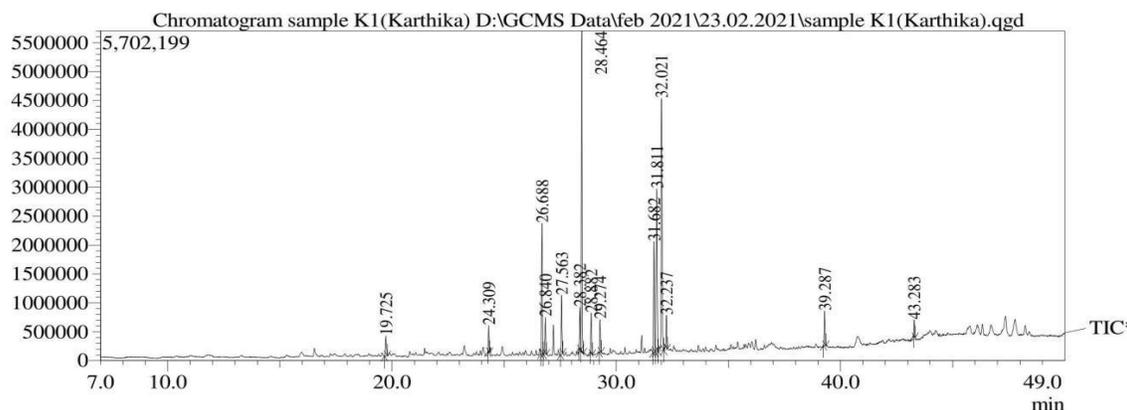
3. RESULTS AND DISCUSSION:

GC-MS analysis of methanolic extract of *S.alternata* resulted in the identification of 15 secondary metabolites. Those compounds identified are Neophytadine, 3,7,11,15-Tetramethyl 2 hexadecen-1ol, Methyl palmitate, Methyl octadeca 9,12 Dienoate, Methyl laurate, Methyl myristate, Hexahydrofarnesyl acetone, 9-octadecenoic acid, Isophytol, 2,5 Dimethyl-4 Hexen 3 ol, Linolenic acid, Phytol, Methyl stearate, 1,2 Benzene dicarboxylic acid and Squalene. Quantitative expression of these secondary metabolites was expressed by considering the area percentage of GC-MS profile (Table-1:Chromatogram- 1).

TABLE-1 GCMS DATA

Sl.NO	NAME OF THE SECONDARY METABOLITES	AREA PERCENTAGE
1	Neophytadine	9.09
2	Hexahydrofarnesyl acetone	2.68
3	Isophytol	2.60
4	Phytol	20.57
5	Squalene	1.02
6	3,7,11,15 tetramethyl 2 hexadecen 1 ol	4.07
7	Methyl palmitate	22.28
8	Methyl laurate	1.80
9	Methyl myristate	1.55
10	9-Octadecenoic acid	2.69
11	2,5 dimethyl 4 hexen 3 ol	2.31
12	Linolenic acid	16.05
13	Methyl stearate	2.77
14	1,2-benzene dicarboxylic acid	2.66
15	Methyl octadeca 9,12 dienoate	7.86

CHROMATOGRAM OF GCMS DATA



Considering the area percentage abundantly occurring component is Methyl palmitate(22%). Second abundantly occurring compound was phytol (20%) followed by Linolenic acid(16%).In addition to the above components the following fatty acid methyl esters were also present as given below.

Methyl octadeca 9,12 dienoate (7.86%), 3,7,11,15 Tetramethyl 2 hexadecen 1-ol (4.07%), Methyl stearate (2.77%), 9-Octadecenoic acid(2.69%), Methyl laurate (1.80%) and Methyl myristate (1.55%).Other than the Fatty acid methyl esters, six terpenoid compounds also were identified in the GC-MS analysis of *S.alternata* leaves. Phytol with area percentage 21% is the abundantly occurring monounsaturated diterpene alcohol. Neophytadine was second abundantly occurring terpenoid (9%) belonging to sesquiterpenoid. Hexahydrofarnesyl acetone content was 2.7% and Isophytol was 2.60%. Least distributed bioactive compound was squalene, a triterpene present only 1.02%.1,2-Benzene dicarboxylic acid(2.66%) was an aromatic dicarboxylic acid present in *S.alternata* other than the terpenoids and fatty acid methyl esters.

Medicinal property of *H.colorata* is known earlier and used for wound healing as a folk medicine. Research publication on medicinal use of this plant reveals the antimicrobial [23] and anti-inflammatory properties[20]. Antioxidant activity has also been mentioned recently[24],[25] . All these investigations are based on experiments with microbes and for test animals subjected to treatment with aqueous and alcoholic extracts of *S.alternata*. However the principles and/or phytochemical roles of the component of the extracts have not yet been delineated.

GCMS study on alcohol extract of *S.alternata* (*H.colorata*) revealed the occurrence of a number of

bioactive components (Table- 1) of which many molecules possess active principles pertaining to the medicinal use for wound healing. Methyl palmitate is abundantly occurring in *S.alternata* and this component is having anti-inflammatory property [26,27]. Antiinflammatory potential of this plant is found to be due to the presence of Neophytadine, 3,7,11,15-Tetramethyl 2 hexadecen 1 ol because these molecules are known to play anti- inflammatory role in accordance with the view[28]. Another bioactive molecule present in *S.alternata* is phytol which is the second abundant component possessing antimicrobial, antinociceptive, antioxidant and immunostimulant activities as opined by Rhy *et al.*,(2011) and Santos *et al.*,(2012)[29,30]. Hexahydrofarnesyl acetone (sesquiterpene), Isophytol are potential molecules characterized by antibacterial and antifungal properties as reported by VenkataRaman *et al.*,(2012). Methyl stearate is present in the leaf extract of *S.alternata* [28]. Ukwubile *et al.*,(2019) stated that methyl stearate isolated from *Melastromatum capitatum* play a vital role in health care systems and according to those authors presence of methyl stearate and fatty acids in the leaves of the plant might be responsible for its biological activity in traditional medicine[31].

Antifungal activity of fatty acid methyl esters of vegetable oils has been demonstrated and evaluated in case studies on fungus [32]. Antioxidant potential was demonstrated by scavenging effect of these fatty acid methyl esters on 2,2-diphenyl 1- picryl hydrazyl (DPPH) radicals.

Antioxidant activities of many secondary metabolite molecules is well known and excellently interpreted in the process of wound healing [33,34]. 3,7,11,15-tetramethyl 2 hexadacen 1 ol, 9-octadecenoic acid, phytol, fatty acid methyl esters and squalene are the bioactive components present in *S.alternata* which

are characterized by antioxidant potential[28,30,32,36]. Antioxidant potential of *S.alternata* extract has been demonstrated by DPPH assay[24,25].

Houghton *et al.*,(2005) reported the involvement of antioxidant components in wound healing process has in *Secamone alzei* and *Spathodea campanulata*[35]. In addition a number of plants that have been reported to show antioxidant potential coming under different families[34]. Squalene is a triterpene which occurs comparatively in very small quantity in *S.alternata* (Table-1). According to Lozano-Granta *et al.*,(2018) squalene is a biochemical intermediate possessing antioxidant activity present in all vegetable oils[37]. Aioi *et al.*,(1995) reported the scavenging role of squalene on ROS produced by oxidative stressors and have a protective role of this molecule in combination with SOD[38]. Squalene is abundant in skin which is the target tissue exposed to environmental stresses leading to oxidative stress and hence squalene plays an antioxidant role in protecting the skin due to the specific role of squalene [36].

Shetty *et al.*,(2007) interpreted that wound healing is the process of repair that follows injury to the skin and initial stages of wound healing involves an acute inflammatory phase[33]. According to those authors, the antioxidant properties of *Ocimum sanctum* is found to be responsible for faster wound healing in rats. Incision wounds in rats resulted in the increased antioxidant enzymes and *Ocimum* extract treatment showed decreased activity indirectly establishing antioxidant potential of *Ocimum sanctum* in wound healing[33].

According to Dissemmond *et al.*,(2002) in the therapy of chronic wounds overproduction of ROS results in the oxidative stress, thereby causing cytotoxicity and delayed wound healing[39]. Antioxidants like SOD, catalase, Glutathione etc hasten the process of wound healing by destroying the ROS .It is known that the wound healing process can be aided by the presence of antioxidants and 36 plants have been reported to possess wound healing and antioxidant activity[34].

As mentioned earlier, besides antibacterial and antifungal properties, antioxidant potential is another important property of squalene[37]. According to those authors, in human body squalene is secreted by sebaceous glands for skin protection and forms 10-15% of lipid on skin surface and on internal organs such as liver. This behaviour of squalene is found to be correlated with the practise of topical application

of leaf juice of *S.alternata* to fresh wounds for immediate healing is consistent with the view of Subramaniam *et al.*,(2001)[20]. Antioxidant property of squalene is also related to the protection by scavenging the ROS produced due to wound- induced stress whereas antimicrobial activity of those components of the leaf juice performs an aseptic condition to enhance wound-healing indicating an antiseptic property of *S.alternata* resulting in immediate healing of wounds.

The antioxidant properties of squalene and use as a topical skin lubricant and protectant can also be correlated with wound healing in consonance with the view of Micera *et al.*,(2020) according to whom squalene possess antioxidant activity and is abundant in skin which is the target tissue exposed to wounds or other incisions[36].

The vital role of antioxidants in the process of wound healing is found to be due to the presence of phytol, squalene, 9-octadecenoic acid, 3,7,11,15 tetramethyl 2 hexadacen 1 ol, methyl palmitate and other fatty acid methyl esters of *S.alternata*. This observation is in accordance with the view of Suntar *et al.*,(2007) according to whom wound healing and antioxidant activity coexist in many plant species of a variety of families[34]. As mentioned earlier, wound healing capacity of *S.alternata* is already known and presumed to involve antimicrobial, anti-inflammatory and antioxidant activities of leaf juice[20]. The phytochemical functions of each bioactive component of leaf extract play a vital role in the process of wound healing which is found to be a concerted activity of molecules present in the leaf extracts. Supplementary role of hexahydrofarnesyl acetone, methyl stearate, linolenic acid and 9-octadecenoic acid compounds in the process of wound healing and its complementary role cannot be ruled out.

4. CONCLUSION:

Bioactive property of secondary metabolites in leaf extract of *Strobilanthes alternata* reveals the antimicrobial, anti-inflammatory and antioxidant activities. The involvement of these molecules in the process of wound healing is a known character of the plant is found to be due to the concerted activity of 15 biomolecules many of them possess antioxidant activity and hence wound healing and antioxidant activity coexist in *S.alternata*.

5. DECLARATION OF COMPETING INTEREST

The authors report no conflicts of interest. The authors alone are responsible for the content and

writing of this article.

6. ACKNOWLEDGMENT

I wish to express my deep sense of gratitude to my teacher, Prof. Dr. Nabeesa Salim, Professor and Head (Retd.), Department of Botany, Calicut University, for the interest in the investigation.

7. REFERENCES:

- Dubey NK, Kumar R, and Tripathi P: Global promotion of herbal medicine: India's opportunity. *Current Science* 2004;86:37-41.
- Rates SMK. Plant sources as drugs. *Toxicol.* Elsevier 2001;39:603-613.
- Alviano DS. and Alviano CS. Plant extracts-search for new alternatives to treat microbial diseases. *Current Pharm. Biotechnology* 2011;10:106-121.
- Ghosh PK. and Gaba A. Phyto-extracts in wound healing. *Journal of pharmacy and pharmaceutical sciences* 2013;16:760-820.
- Ghuman S. Ncube B, Finnie JF, McGaw LJ, Njoya EM, Coopoosamy RM. and Staden JV. Antioxidant, Anti-inflammatory and wound healing properties of medicinal plant extracts used to treat wounds and dermatological disorders. *South African Journal of Botany* 2019; 126:232-240.
- Phrompittayarat W, Jetiyanon K, Wittaya Areekul S. and Putaloun W. Influence of seasons, different plant parts, and plant growth stages on Saponin quantity and distribution in *Bacopa monnieri*. *Songklanakar J.Sci. Technol* 2011;33:193-199.
- Singh D and Aeri V. Phytochemical and pharmacological potential of *Acanthus ilicifolius*. *J.Pharm Bioallied Sci* 2013;5:17-20.
- Kuppuswamy KM, Jounalagadda B and Arockiasamy S. GC-MS analysis of chloroform extract of *Croton bonplandianum*. *Int. J. Pharma. Bio. Sc.* 2013; 4:613-617.
- Abid S and Touqueer S. Antimicrobial and antioxidant activity of *Breynia disticha* and *Vernonia elaeagnifolia*. *J.App.Pharm* 2015;7:178-182.
- Tamboli FA, Rangari VD, Killedar SG, Jadhar S U, Ghatage TS and Kore VP. Comparative phytochemical evaluation of natural micropropagated plants of *Bacopa monnieri* (L.) and Marmara. *Pharm J.* 2018 ; 22:66-73.
- Yu FR, Lian XZ, Guo HY, McGuire PM, Li RD, Wang R and Yu F H. Isolation and characterization of methyl esters and derivatives from *Euphorbia kansui* (Euphorbeaceae) and their inhibitory effects on the human SCG-7901 cells. *J.Pharm.Pharm Sci* 2005;8:528-535.
- Sermakkani N and Thangapandian V. GC-MS analysis of *Cassia italic* leaf methanol extract. *Asian J of Pharmaceutical and clinical research.* 2011;5:90-94.
- Ryu KR, Choi JY, Chung S and Kim DH. Anti scratching behavioural effect of the essential oil and phytol isolated from *Artemisia princeps* Pamp. in mice. *Planta Med* 2011;77:22-26.
- Al-Owaisi M, Al-Hadiwi N and Khan SA. GCMS analysis, determination of total phenolics, flavonoid content and free radical scavenging activities of various crude extracts of *Moringa peregrina* (Forssk.) Flori leaves *Asian Pacific Journal of Tropical Biomedicine* 2014;4:964-970.
- Igwe KK, Nwankwo PO, Otuokere IE, Ijioma S N and Amaku FM. GCMS analysis of phytocomponents in the methanolic extract of *Moringa oleifera* leaves. *Journal of Research in pharmaceutical science* 2015;2:01-06.
- Igwe KK, Nwankudu ON, Ijoma SN, Madubuike AJ and Achi NK. Screening of secondary metabolites in *Huru crepitans* bark ethanol extract using GC-MS analysis : A preliminary study approach. *Journal of Science and Technology Advances* 2016;1:64-71.
- Mercurio I, Melai P, Capan D, Ceraso G, Carlini L and Bacci M. GCMS analysis of morning glory seeds freely in commerce: Can they be considered "herbal highs"? *Egyptian Journal of Forensic sciences* 2017; 7:1-6.
- Balogun OS, Ajayi OS and Adeleke AJ . Hexahydrofarnesyl acetone rich extractives from *Hildegardia barteri*. *Journal of herbs, spices and medicinal plants* 2017;10:1049-6475.
- Roy SJ , Baruah PS, Lahikar L, Gurung L , Saikia D and Tanti B. Phytochemical analysis and antioxidant activities of *Homolomena aromatic* Schott. *Journal of Pharmacognosy and Phytochemistry* 2019;8: 1379-1385.
- Subramoniam A, Evans DA, Rajasekharan S and Nair GS. Effect of *Hemigraphis colorata* (Blume) H.G. Hallier leaf on wound healing and

- inflammation in mice. *Int.Journal of Pharmacology* 2001;33:283-285.
21. Bhargavi CHS, Kumar NVSPP and Babu VR. Ancient and modern view of wound healing: Therapeutic Treatments.RJPBCS 2011;2:474-479.
 22. Pawar RS and Toppo FA.Plants that heal wounds:A review. *Herba polonica* 2012;58.
 23. Anitha VT,Antonisamy JM and Jeeva S. Antibacterial studies on *Hemigraphis colorata* (Blume) H.G.Hallier and *Elephantopus scaber* L. *Asian Pac J Trop Med* 2012; 5:52-57.
 24. Megha MA, Unnma U, Rameshpathy M, Karikalan K,Vickram S,KumaRSV,Sridharan B. Formulation of nano-encapsulated poly herbal ointment for anti-inflammation.*Der Pharmacia Lettre* 2013;5:164- 170.
 25. Akhil, P. and Prabhu, P. (2013). Evaluation of antioxidant, anti-inflammatory and cytotoxicity potential of *Hemigraphis colorata*. *IJPSR*. 2013;4:3477-3483.
 26. Goswami U and Nazarine F. Bioactivity of methyl palmitate obtained from mangrove plant *Salvadora persica* L. US Patent 2003;6: 638,546.
 27. Wang YN, Wang HX, Shen ZJ, Zhao LL, Clarke SR, Sun JH, Du YY and Shi GL. Methyl palmitate, an acaricidal compound occurring in green walnut husks. *Jornal of economic entomology* 2009; 102: 196-202.
 28. VenketaRaman B, Samuel LA , Pardhasaradhi M, NarashimhaRao B, Nagavamskrishna A, Sudhakar M and Radhakrishnan TM. Antibacterial,antioxidant activity and GCMS analysis of *Eupatorium odoratum*. *Asian J. Pharm Clin Res* 2012; 5:99-106.
 29. Ryu KR,Choi JY,Chung S and Kim DH. Anti scratching behavioural effect of the essential oil and phytol isolated from *Artemisia princeps* Pamp. in mice.*Planta Med* 2011;77:22-26.
 30. Santos CCMP, Salvadori MS, Mota VG, Costa LM and Almeida AACO. Antinociceptive and antioxidant activities of phytol *invivo* and *invitro* models. *Neurosci J*. 2013;Article ID 949452.
 31. Ukwubile CA, Ahmed A, Katsayal UA, Ya UJ and Mejida S..GCMS analysis of bioactive compounds from *Melastomastrum capitatum*(Vahl) Fern leaf methanol extract:an anticancer plant. *Scientific African*. 2019;3:e0059.
 32. Pinto MEA, Araujo SG, Morais MI, Sa,NP, Lima CM, Rosa CA, Siqueria EP, Johann S and Lima LARS. Antifungal and antioxidant activity of fatty acid methyl esters from vegetable oils. *Anal. da Academia Brasileira de ciencias* 2017; 89:1671-1681.
 33. Shetty S, Udupa S and Udupa L. Evaluation of antioxidant and wound healing effects of alcoholic and aqueous extract of *Ocimum sanctum* Linn in rats. *Advance Access Publication* 2007; 5:95-101.
 34. Suntar I, Akkol EK, Nahar L and Sarker SD. Wound healing and antioxidant properties: do they coexist in plants ? *Free Rad.Antiox* 2012;2:1-7.
 35. Houghton PJ, Hylands PJ, Mensah AY, Hensel A and Deters AM. Invitro tests and ethnopharmacological investigations. Wound healing as an example. *Journal of Ethanopharmacology* 2005;100:100-107.
 36. Mishra M, Botto, Geddo F, Antoniotti S, Berteau CM, Levi R, Gallo MP and Querio G. Squalene: More than a stem toward sterols. *Antioxidants* 2020; 9:688.
 37. Lozano-Grande M, Gorinstein S, Espitia-Rangel E, Davila-Ortiz G and Martinez-Ayala AI. Plant sources, extraction methods and uses of squalene. *International journal of Agronomy* 2018; ID 1829160:13.
 38. Aioi A, Shimizu T, and Kuriyama K. Effect of squalene on superoxide anion generation induced by a skin irritant, lauroylsarcosine. *Int J Pharm* 1995; 113:159-164.
 39. Dissemond J, Goos M and Wagner SN. The role of oxidative stress in the pathogenesis and therapy of chronic wounds. *Hautarzt* 2002; 53:718-23.