



CODEN [USA]: IAJPBB

ISSN : 2349-7750

## INDO AMERICAN JOURNAL OF PHARMACEUTICAL SCIENCES

SJIF Impact Factor: 7.187

<https://doi.org/10.5281/zenodo.7277652>Available online at: <http://www.iajps.com>

Review Article

### INDICATION AND CRITERIA OF PERIPHERAL BLOOD SMEAR

**Deemah Hasin Algurashi, Raghda AbdulRahman AbdulMajeed, Samaher Adnan Bulkhi,  
Jehad Abdulrashid Bifari, Omar Seraj Bakhsh, Khalid Othman Hawsawi, Manal  
Abdulhalim Mohammed Shiqdar, Maha Adnan Sabbagh**

<b>Article Received:</b> September 2022	<b>Accepted:</b> September 2022	<b>Published:</b> October 2022
---	---------------------------------	--------------------------------

**Abstract:**

*A peripheral blood smear review is a standard test used to diagnose both hematologic and non-hematologic illnesses. An expert's evaluation of a peripheral smear allows for a broad examination of blood cell morphology and may identify metabolic, nutritional, genetic, and inflammatory problems, as well as hemolysis, blood-borne parasites, and neoplasia. Literature review among previous published studies up to November, 2021, carried out to reach the concept of indication for peripheral blood smear. In contrast, smears requested to evaluate anemia and thrombocytopenia, the most common reasons for peripheral smear review requests, had lower rates of potential clinical value, aside from confirming the absence of significant findings.*

**Corresponding author:**

**Deemah Hasin Algurashi,**

QR code



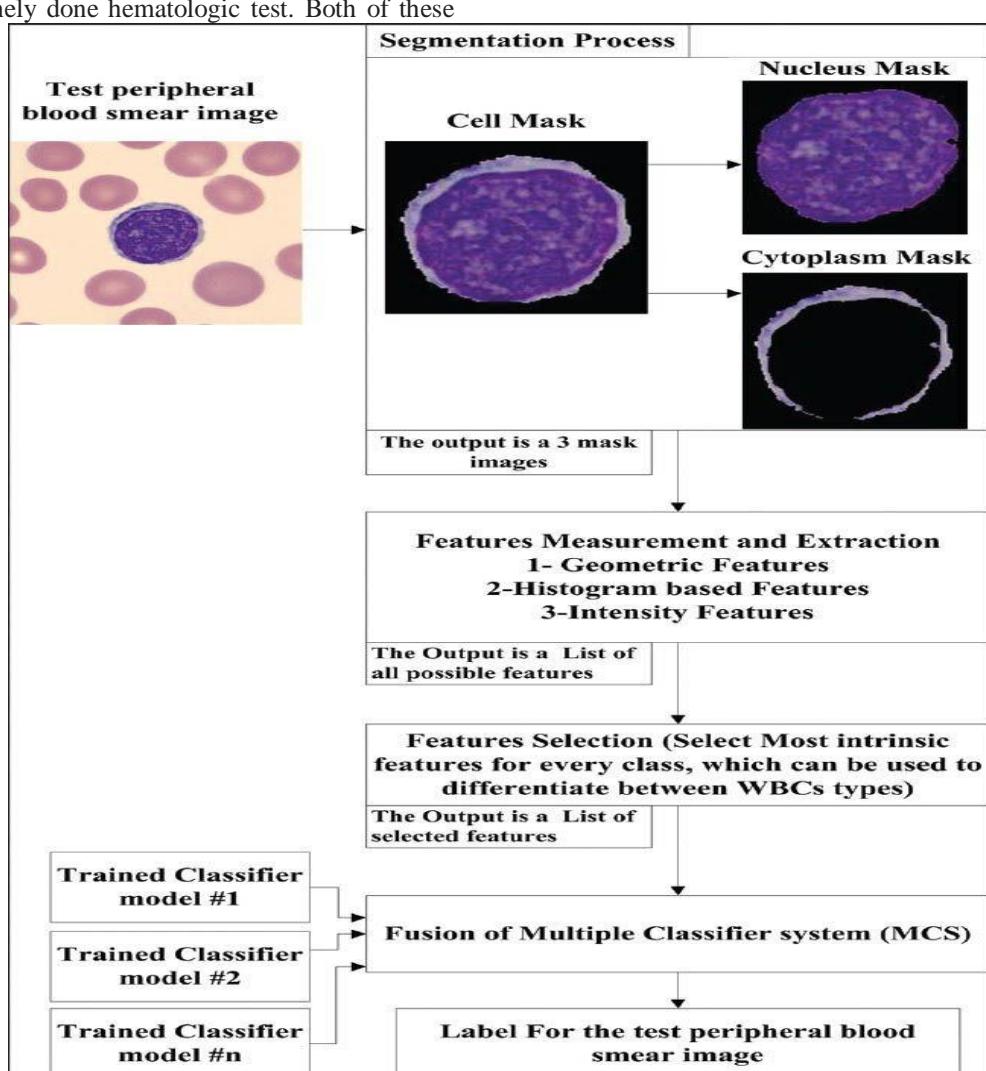
**Please cite this article in Deemah Hasin Algurashi *et al*, Indication And Criteria Of Peripheral Blood Smear., Indo Am. J. P. Sci, 2022; 09(10).**

## INTRODUCTION:

A blood smear review (BSR) is described as a meticulous microscopic examination of a well-prepared and stained smear of peripheral blood with the goal of identifying morphological alterations important to patient diagnosis and monitoring [1]. It is also regarded as an internal quality control tool for evaluating the parameters offered by hematology analyzers. BSR is one of the most time-consuming processes in hematology laboratories, requiring great technical expertise to prevent errors inherent in the subjectivity of MBSR, such as manual differential leukocyte counts (MDLC) [2,3].

A complete blood count, often known as a CBC or, less frequently, a Hemogram, is the most common test performed in a clinical hematology laboratory. The differential leukocyte count, or DIFF, is the second most routinely done hematologic test. Both of these

tests can be performed fairly reliably, efficiently, and cost-effectively using currently available automated hematology analyzers [4,5]. A qualified laboratory professional's microscopic evaluation of a correctly prepared and well-stained blood smear is, nevertheless, important and clinically valuable in a variety of contexts and for a variety of reasons [6,7]. Screening prepared blood films is time-consuming, tedious, and sensitive to inter- and intra-observer variance. This has consequences for both laboratory resources and diagnostic accuracy. As a result, many studies have addressed issues with microscopic image analysis of peripheral blood smears, and establishing image analysis methods for computer-assisted interpretations of peripheral blood smears is critical. (Figure 1) depicts the fundamental workflow processes in a peripheral blood smear image analysis system [8,9].



**Figure 1:** Workflow of peripheral blood smear image analysis, starting from image segmentation and features extraction

**DISCUSSION:**

A peripheral blood smear review is a standard test used to diagnose both hematologic and non-hematologic illnesses. An expert's evaluation of a peripheral smear allows for a broad examination of blood cell morphology, which may indicate metabolic, nutritional, genetic, and inflammatory problems, as well as hemolysis, blood borne parasites, and neoplasia [10]. Smear examination is frequently used as a reflex test to examine abnormalities detected by automated tests such as complete blood count with differential (CBC-D) [11]. Clinical concern for hemolysis, hematolymphoid neoplasms, and other disorders marked by abnormalities in blood cell morphology are also indications [12]. Despite the growth of automated laboratory tests in recent decades, smear review remains an agnostic test with the ability to reveal information beyond what automated testing alone can identify [11]. Furthermore, sample collection provides little risk to the patient, and the smear itself is simple and rapid to create.

Interestingly, in some situations, the reason for the smear was not directly related to the potentially significant discovery, implying that the discovery was unanticipated. These smear reviews were a subset of all studies with potential added clinical value; a list of smears with surprising findings is included in the report (**Table1**). The evaluation of peripheral blood smears is widely acknowledged as an important stage

in the workup of many clinical conditions. However, the test predates the automated hematological and chemistry assays that are currently ubiquitous in resource-rich medical institutes. Modern hematology analyzers, for example, can detect platelet clumping as well as assess numerous RBC properties such as mean corpuscular volume, mean corpuscular hemoglobin concentration, and red cell dispersion width. These metrics analyze RBC morphology and may detect pseudo-thrombocytopenia, obviating the necessity for manual examination in some circumstances [12].

The list of criteria for smear review is usually developed by individual laboratories with input from pathologist(s), clinicians, and the hematology supervisory staff, and may be updated periodically as deemed appropriate. Although, clinical significance of the abnormal CBC and DIFF findings is the major determining factor in deciding which blood smears need review, several other factors may also influence such a decision. These factors may include patient population served, clinicians' concerns pertaining to specific patient populations, training and experience of blood smear examiners(s) and reviewer(s), workload of the laboratory and the reviewer(s), initial vs. follow-up smears, QC/quality assurance (QA) consideration, and teaching/educational considerations. Published criteria may be used by individual laboratories as a starting point in the process of developing their own set of criteria [13,14].

**Table 1:** Findings among individual smears requested for select indications

Indication	Findings seen among individual smears requested for the specified indication
Leukocytosis	<ul style="list-style-type: none"> <li>• Atypical / variant lymphoid cells</li> <li>• Findings consistent with a hemoglobinopathy</li> <li>• Leukemia or blasts</li> <li>• Rouleaux</li> </ul>
Anemia	<ul style="list-style-type: none"> <li>• Atypical/variant lymphoid cells</li> <li>• Dysplasia</li> <li>• Findings consistent with a hemoglobinopathy</li> <li>• Hemolysis</li> <li>• Leukemia or blasts</li> <li>• Other</li> <li>• Platelet abnormality</li> <li>• Rouleaux</li> </ul>
Leukopenia	<ul style="list-style-type: none"> <li>• Atypical/variant lymphoid cells</li> <li>• Dysplasia</li> <li>• Findings consistent with a hemoglobinopathy</li> <li>• Hemolysis</li> <li>• Leukemia or blasts</li> <li>• Platelet abnormality</li> </ul>

Thrombocytopenia	<ul style="list-style-type: none"> <li>• Atypical/variant lymphoid cells</li> <li>• Dysplasia</li> <li>• Hemolysis</li> <li>• Leukemia or blasts</li> <li>• Platelet abnormality</li> <li>• Rouleaux</li> </ul>
Hemolysis	<ul style="list-style-type: none"> <li>• Dysplasia</li> <li>• Findings consistent with a hemoglobinopathy</li> <li>• Hemolysis</li> <li>• Leukemia or blasts</li> <li>• Other</li> <li>• Platelet abnormality</li> <li>• Rouleaux</li> </ul>

Several of the reports describing the detection of yeast in blood smears suggested microscopic review as a method for early detection of yeast fungemia. The usefulness of blood smear review for this purpose ultimately depends on the frequency of very high yeast levels in clinical samples. Therefore, it is important to ascertain how frequently yeast levels of  $10^5$  and higher (the concentration we found was necessary to detect yeast in a blood smear) occur in clinical samples. In order to address this question, we reviewed the relevant literature. Several groups have used quantitative cultures to measure the concentration of yeast in blood samples from fungemic patients [15]. Recent investigations have applied molecular methods to the quantification of *Candida* in blood samples, and these methods are not subject to this limitation. For example, Maaroufi et al [16,17] described a polymerase chain reaction-based assay to quantify *C albicans* in blood samples from patients with clinically proven or suspected systemic *Candida* infections. They found a wide range of *C albicans* loads among 11 positive samples tested, extending from 5 to 100475 CFU/mL [17].

### CONCLUSION:

The review of peripheral blood smears is a time-consuming, under-reimbursed investigation with uncertain therapeutic relevance. According to this multi-institutional study, smear review ordering processes vary, but result in a small number of smear reviews with potential clinical benefit beyond automated laboratory testing alone. The indications with the highest potential clinical value rates (WBC number, WBC morphology, and assessment for hematolymphoid malignancy). Smears requested to check anemia and thrombocytopenia, the two most prevalent causes for peripheral smear evaluation, had lower rates of possible clinical usefulness, aside from verifying the absence of noteworthy abnormalities.

### REFERENCES:

1. Gulati G.L., Alomari M., Kocher W., Schwarting R. Criteria for blood smear review. *Lab Med*. 2002;33(5):374–377.
2. Pierre R.V. Peripheral blood film review: the demise of the eyecount leukocyte differential. *Clin Lab Med*. 2002;22(1):279–297.
3. Pierre R.V. Red cell morphology and the peripheral blood film. *Clin Lab Med*. 2002;22(1):25–61.
4. Carr J, Geesaman S, Czader M. Performance evaluation of the new Unicel DxH 800 Coulter Cellular Analysis system in a large hospital setting. *Lab Med*. 2012;43:157–163.
5. Kang SH, Kim HK, Ham CK, Lee DS, Cho HI. Comparison of four hematology analyzers, Cell-DYN Sapphire, Advia 120, Coulter LH750, and Sysmex XE-2100, in terms of clinical usefulness. *Int J Lab Hematol*. 2008;30:480–486.
6. Peterson P, McNeill S, Gulati G. Cellular morphologic analysis of peripheral blood. In: Kottke-Marchant K, Davis BH, editors. *Laboratory hematology practice*. Chichester: Wiley-Blackwell; 2012. pp. 10–25.
7. Bain BJ. Diagnosis from the blood smear. *N Engl J Med*. 2005;353:498–507.
8. Mohammed EA, Mohamed MM, Naugler C, Far BH. 26th Annual IEEE Canadian Conference on, IEEE; 2013. Chronic lymphocytic leukemia cell segmentation from microscopic blood images using watershed algorithm and optimal thresholding. Electrical and Computer Engineering (CCECE), 2013.
9. Guo N, Zeng L, Wu Q. A method based on multispectral imaging technique for white blood cell segmentation. *Comput Biol Med*. 2007;37:70–6.

10. Rosenthal D. Evaluation of the peripheral blood smear. Waltham: UpToDate, Inc; 1993. Accessed 15 Sept 2019.
11. Barnes PW, McFadden SL, Machin SJ, Simson E. The international consensus group for hematology review: suggested criteria for action following automated CBC and WBC differential analysis. *Lab Hematol.* 2005;11(2):83–90.
12. Bain BJ. Diagnosis from the blood smear. *N Engl J Med.* 2005;353(5):498–507.
13. Javidian P, Garshelis L, Peterson P. Pathologist review of the peripheral film. A mandatory quality assurance activity. *Clin Lab Med.* 1993;13:853–861.
14. Peterson P, Blomberg DJ, Rabinovitch A, Cornbleet PJ. Physician review of the peripheral blood smear: when and why—an opinion. *Lab Hematol.* 2001;7:175–179.
15. Henry, N. K., C. A. McLimans, A. J. Wright, R. L. Thompson, W. R. Wilson, and J. A. I. Washington. Microbiological and clinical evaluation of the isolator lysis-centrifugation blood culture tube. *J Clin Microbiol* 1983. 17:864–869.
16. Maaroufi, Y., N. Ahariz, M. Husson, and F. Crokaert. Comparison of different methods of isolation of DNA of commonly encountered *Candida* species and its quantitation by using a real-time PCR-based assay. *J Clin Microbiol* 2004. 42:3159–3163.
17. Maaroufi, Y., C. Heymans, and J. M. De Bruyne. et al. Rapid detection of *Candida albicans* in clinical blood samples by using a TaqMan-based PCR assay. *J Clin Microbiol* 2003. 41:3293–3298.